

BILATEST PCR Cleanup Kit



100 Extractions

Product description

The BILATEST PCR Cleanup Kit is designed for the simple and fast purification of PCR products.

This kit contains enough materials for 100 x 50 µl PCR product purifications and is scalable for other sample volumes.

Included reagents

Reagent	Volume
(1) Magnetic Beads	1.2 ml
(2) Binding Buffer	14 ml
(3) Washing Buffer A	30 ml
(4) Washing Buffer B	60 ml
(5) Elution Buffer	5 ml

Required Materials

96% or 100% Ethanol

This kit is optimized for use with BILATEC Magnetic Separators (e.g. BILATEST magnetic separator M 12+12 for 1.5ml tubes, Order-No. 210141).

Storage Conditions and Safety Information

This kit may be stored at room temperature (15 – 25°C) and is stable for at least 1 year following delivery. The kit buffers contain irritant substances. Take appropriate laboratory safety measures and wear gloves when handling.

Samples and Protocol Adjustments

Included in this kit is one **Elution Buffer (5)** (10 mM Tris-HCl, pH 8.0), TE buffer pH 8.0 can also be used without any protocol adjustments. Water may also be used, in this case we recommend an elution time of 10 – 15 minutes at 55°C to ensure a high yield of purified product.

This protocol is completely scalable for other PCR product volumes.

Before you Start

Add 48 ml 96% or 100% Ethanol to the **Washing Buffer B (4)** bottle.

Purification Protocol for 50 µl PCR Products

1. Place the **50 µl PCR product** in a 1.5 ml microfuge tube. Add **12 µl Magnetic Beads (1)** and **138 µl Binding Buffer (2)**. Be sure to resuspend the Magnetic Beads before dispensation; if they have been standing for a long period of time it may help to vortex them briefly before removal.
2. Mix with 4 – 5 pipetting strokes and incubate **5 minutes at room temperature**.
3. Following incubation, separate the **Magnetic Beads (1)** against the side of the tube. Wait 30 seconds or until all the beads have been attracted to the magnet. Pipette off the supernatant and then remove the tube from the magnet.
4. Add **300 µl Washing Buffer A (3)** to the tube and thoroughly resuspend the beads in the washing buffer by pipetting the bead pellet up and down 4 times.
5. Separate all of the **Magnetic Beads (1)** against the side of the tube and then discard the supernatant.
6. Remove tube from the magnet position and repeat the washing procedure (steps 4 and 5) using **300 µl Washing Buffer B (4)**.
7. Repeat washing step 6 using **300 µl Wash Buffer B (4)**.
8. Incubate the tube 10 minutes at room temperature with the particles held against the magnet and the lid open in order to allow the remaining traces of alcohol to evaporate.
9. Add **50 µl** (or another suitable volume) of **Elution Buffer (5)** to the tube and resuspend the Bead/PCR Product complex by pipetting.
10. Incubate the suspension for **5 minutes at 55°C**, with occasional agitation, to facilitate complete elution of the purified PCR product.
11. Following elution, separate the **Magnetic Beads (1)** against the side of the tube for 2 minutes to remove all of the beads from suspension.
12. Transfer the eluate containing the purified PCR Product to a clean tube. Store under appropriate conditions or use for subsequent applications.